OptiPrep™ Reference List RV05

GROUP V VIRUSES

- **Viruses are listed alphabetically within the Baltimore scheme: Family, Genus and Species. Publications are listed alphabetically by first author and, where necessary, references are further divided according to research topic**
- **Multiple entries from the same first author are listed chronologically.**
- **For a detailed methodology of Group V viruses see OptiPrep™ Application Sheets V23-V28. V06 is a methodological review of OptiPrep™ technology.**

1 *Arenaviridae*

Arenavirus

Junin virus

Chou, Y-y., Cuevas, C., Carocci, M., Stubbs, S.H., Ma, M., Cureton, D.K., Luke Chao, L., Evesson, F. et al (2016) *Identification and characterization of a novel broad-spectrum virus entry inhibitor* J. Virol., **90**, 4494-4510

Gaudin, R. and Barteneva, N.S. (2015) *Sorting of small infectious virus particles by flow virometry reveals distinct infectivity profiles* Nat. Commun, **6**: 6022

Gaudin, R. and Kirchhausen, T. (2015) *Superinfection exclusion is absent during acute Junin virus infection of Vero and A549 cells* Sci. Rep., **5**: 15990

Lassa virus

Baird, N.L., York, J. and Nunberg, J.H. (2012) *Arenavirus infection induces discrete cytosolic structures for RNA replication* J. Virol., **86**, 11301-11310

Eichler, R., Lenz, O., Strecker, T. and Garten, W. (2003) *Signal peptide of Lassa virus glycoprotein GP-C exhibits an unusual length* FEBS Lett., 538, 203-206

Lenz, O., Ter Meulen, J., Feldmann, H., Klenk, H-D. and Garten, W. (2000) *Identification of a novel consensus sequence at the cleavage site of the Lassa virus glycoprotein* J. Virol., 74, 11418-11421

Strecker, T., Eichler, R., ter Meulen, J., Weissenhorn, W., Klenk, H.D., Garten, W. and Lenz, O. (2003) *Lassa virus Z protein is a matrix protein sufficient for the release of virus-like particles* J. Virol., **77**, 10700-10705

Ziegler, C.M., Eisenhauer, P., Bruce, E.A., Weir, M.E., King, B.R., Klaus, J.P., Krementsov, D.N. et al (2016) *The lymphocytic choriomeningitis virus matrix protein PPXY late domain drives the production of defective interfering particles* PLoS Pathog., **12**: e1005501

2 *Bunyaviridae*

Bunyamweravirus

Ariza, A., Tanner, S.J., Walter, C.T., Dent, K.C., Shepherd, D.A., Wu, W., Matthews, S.V., Hiscox, J.A., Green, T.J. et al (2013) *Nucleocapsid protein structures from orthobunyaviruses reveal insight into ribonucleoprotein architecture and RNA polymerization* Nucleic Acids Res., **41**, 5912–5926

Cabezas, P. and Risco, C. (2006) *Studying cellular architecture in three dimensions with improved resolution: Ta replicas revisited* Cell Biol. Int., **30**, 747-754

Hover, S., Foster, B., Fontana, J., Kohl, A., Goldstein, S.A.N., Barr, J.N. and Mankouri, J. (2018) *Bunyavirus requirement for endosomal K^+ reveals new roles of cellular ion channels during infection* PLoS Pathog **14**: e1006845

Novoa, R.R., Calderita, G., Cabezas, P., Elliott, R.M. and Risco, C. (2005) *Key Golgi factors for structural and functional maturation of bunyamwera virus* J. Virol., **79**, 10852-10863

Hantavirus

Bisoffi, M., Hjelle, B., Brown, D.C., Branch, D.W., Edwards, T.L., Brozik, S.M., Bondu-Hawkins, V.S. and Larson, R.S. (2008) *Detection of viral bioagents using a shear horizontal surface acoustic wave biosensor* Biosens. Bioelectron., **23**, 1397-1403

Buranda, T., Wu, Y., Perez, D., Jett, S.D., BonduHawkins, V., Ye, C., Edwards, B., Hall, P., Larson, R.S., Lopez, G.P., Sklar, L.A. and Hjelle, B. (2010) *Recognition of decay accelerating factor and avb3 by inactivated*

hantaviruses: Toward the development of high-throughput screening flow cytometry assays Anal. Biochem., **402**, 151–160
Guo, Y., Wang, W., Sun, Y., Ma, C., Wang, X., Wang, X., Liu, P., Shen, S. et al (2016) *Crystal structure of the core region of hantavirus nucleocapsid protein reveals the mechanism for ribonucleoprotein complex formation* J. Virol., **90**, 1048-1061
Hall, P.R., Hjelle, B., Brown, D.C., Ye, C., Bondu-Hawkins, V., Kilpatrick, K.A. and Larson, R.S. (2008) Multivalent presentation of anti-hantavirus peptides on nanoparticles enhances infection blockade Antimicrob. Agents Chemother., 52, 2079-2088
Huiskonen, J.T., Hepojoki, J., Laurinmäki, P., Vaheri, A., Lankinen, H., Butcher, S.J. and Grünewald, K. (2010) *Electron cryotomography of Tula hantavirus suggests a unique assembly paradigm for enveloped viruses* J. Virol., **84**, 4889–4897
Li, S., Rissanen, I., Zeltina, A., Hepojoki, J., Raghwani, J., Harlos, K., Pybus, O.G., Huiskonen, J.T. and Bowden, T.A. (2016) *A molecular-level account of the antigenic hantaviral surface* Cell Rep., **15**, 959–967
Prescott, J.B., Hall, P.R., Bondu-Hawkins, V.S., Ye, C. and Hjelle, B. (2007) *Early innate immune responses to Sin Nombre Hantavirus occur independently of IFN regulatory factor 3, characterized pattern recognition receptors and viral entry* J. Immunol., **179**, 1796-1802

Nairoviridae

Surtees, R., Dowall, S.D., Shaw, A., Armstrong, S., Hewson, R., Carroll, M.W., Mankouri, J., Edwards, T.A., Hiscox, J.A. and Barr. J.N. (2016) *Heat shock protein 70 family members interact with Crimean-Congo hemorrhagic fever virus and Hazara virus nucleocapsid proteins and perform a functional role in the nairovirus replication cycle* J. Virol., **90**, 9305-9326
Wang, X., Li, B., Guo, Y., Shen, S., Zhao, L., Zhang, P., Sun, Y., Sui, S-F., Deng, F. and Lou, Z. (2016) *Molecular basis for the formation of ribonucleoprotein complex of Crimean-Congo hemorrhagic fever virus* J. Struct. Biol., **196**, 455–465

Phlebovirus

Freiberg, A.N., Sherman, M.B., Morais, M.C., Holbrook, M.R. and Watowich, S.J. (2008) *Three-dimensional organization of Rift Valley fever virus revealed by cryoelectron tomography* J. Virol., **82**, 10341-10348
Mbewana, S., Myers, A.E., Rybicki, E.P. (2019) *Chimaeric rift valley fever virus-like particle vaccine candidate production in Nicotiana benthamiana* Biotechnol. J., **14**: 1800238
Weingart, H.M., Zhang, S., Marszal, P., McGreevy, A., Burton, L. and Wilson, W.C. (2014) *Rift valley fever virus incorporates the 78 kDa glycoprotein into virions matured in mosquito C6/36 cells* PLoS One, **9**: e87385
Wolf, M.C., Freiberg, A.N., Zhang, T., Akyol-Ataman, Z., Grock, A., Hong, P.W., Li, J., Watson, N.F., et al (2010) *A broad-spectrum antiviral targeting entry of enveloped viruses* Proc. Natl. Acad. Sci. USA, **107**, 3157–3162

3 *Deltavirus*

Hepatitis D

Perez-Vargas, J., Amirache, F., Boson, B., Mialon, C., Freitas, N., Sureau, C., Fusil, F. and Cosset, F-L. (2019) *Enveloped viruses distinct from HBV induce dissemination of hepatitis D virus in vivo* Nature Comm., **10**: 2098
Verrier, E.R., Colpitts, C.C., Bach, C., Heydmann, L., Weiss, A., Renaud, M., Durand, S.C., Habersetzer, F., Durante, D. et al (2016) *A targeted functional RNA interference screen uncovers glypican 5 as an entry factor for hepatitis B and D viruses* Hepatology, **36**, 35-48

4 *Filoviridae*

Ebola virus

Gélinas, J-F., Azizi, H., Kiesslich, S., Lanthier, S., Perdersen, J., Chahal, P.S., Ansorge, S., Kobinger, G. et al (2019) *Production of rVSV-ZEBOV in serum-free suspension culture of HEK 293SF cells* Vaccine, **37**, 6624–6632
Huang, Y., Xu, L., Sun, Y. and Nabel, G.J. (2002) *The assembly of Ebola virus nucleocapsid requires virion-associated proteins 35 and 24 and posttranslational modification of nucleoprotein* Mol. Cell, **10**, 307-316
Kim, J-O., Chakrabarti, B.K., Guha-Niyogi, A., Louder, M.K., Mascola, J.R., Ganesh, L. and Nabel, G.J. (2007) *Lysis of human immunodeficiency virus type 1 by a specific secreted human phospholipase A2* J. Virol., **81**, 1441-1450
Pastor, A.R., González-Domínguez, G., Díaz-Salinas, M.A., Ramírez, O.T. and Palomares, L.A. (2019) *Defining the multiplicity and time of infection for the production of Zaire Ebola virus-like particles in the insect cell-baculovirus expression system* Vaccine **37**, 6962–6969

5 *Orthomyxoviridae*

Influenza virus

Bungener, L., Serre, K., Bijl, L., Leserman, L., Wilschut, J., Daemen, T. and Machy, P. (2002) *Virosome-mediated delivery of protein antigens to dendritic cells* Vaccine, **20**, 2287-2295

Bungener, L., Huckreide, A., de Mare, A., de Vries-Idema, J., Wilschut, J. and Daemen, T. (2005) *Virosome-mediated delivery of protein antigens in vivo: efficient induction of class I MHC-restricted cytotoxic T lymphocyte activity* Vaccine, **23**, 1232-1241

Chlanda, P., Mekhedov, E., Waters, H., Sodt, A., Schwartz, C., Nair, V., Blank, P.S. and Zimmerberg, J. (2017) *Palmitoylation contributes to membrane curvature in influenza A virus assembly and hemagglutinin-mediated membrane fusion* J. Virol. **91**, e00947-17

Chou, Y-Y., Vafabakhsh, R., Doganay, S., Gao, Q., Ha, T. and Palese (2012) *One influenza virus particle packages eight unique viral RNAs as shown by FISH analysis* Proc. Natl. Acad. Sci. USA, **109,** 9101–9106

Das, D.K., Govindan, R., Nikić-Spiegel, I., Krammer, F., Lemke, E.A. and Munro, J.B. (2018) *Direct visualization of the conformational dynamics of single influenza hemagglutinin trimers* Cell, **174**, 926–937

Galarza, J.M., Latham, T. and Cupo, A. (2005) *Virus-like particle (VLP) vaccine conferred complete protection against a lethal influenza virus challenge* Viral Immunol., **18**, 244-251

Galarza, J.M., Latham, T. and Cupo, A. (2005) *Virus like particle vaccine conferred complete protection against a lethal influenza virus challenge* Viral Immunol., **18**, 365-372

Herbert, A.S., Heffron, L., Sundick, R. and Roberts, P.C. (2009) *Incorporation of membrane-bound, mammalian-derived immunomodulatory proteins into influenza whole virus vaccines boosts immunogenicity and protection against lethal challenge* Virol. J., **6**:42

Hutchinson, E.C., Charles, P.D., Hester, S.S., Thomas, B., Trudgian, D., Martınez-Alonso, M. and Fodor, E. (2014) *Conserved and host-specific features of influenza virion architecture* Nat. Commun., **5**: 4816

Hutchinson, E.C. and Stegmann, M. (2018) *Purification and proteomics of influenza virions* In Influenza Virus Methods and Protocols, Meth. Mol. Biol., **vol. 1836** (ed. Yamauchi, Y.) Springer Science+Business Media New York LLC, pp 89-120

Khan, T., Heffron, C.L., High, K.P. and Roberts, P.C. (2014) *Tailored vaccines targeting the elderly using whole inactivated influenza vaccines bearing cytokine immunomodulators* J. Interferon Cytokine Res., **34**,129-139

Latham, T. and Galarza, J.M. (2001) *Formation of wild-type and chimeric influenza virus-like particles following simultaneous expression of only four structural proteins* J. Virol., **75**, 6154-6165

LeBouder, F., Morello, E., Rimmelzwaan, G.F., Bosse, F., Pechoux, C., Delmas, B. and Riteau, B. (2008) *Annexin II incorporated into influenza virus particles supports virus replication by converting plasminogen into plasmin* J. Virol., **82**, 6820-6828

Le Rua, A., Jacob, D., Transfiguracion, J., Ansorge, S., Henry, O. and Kamena, A.A. (2010) *Scalable production of influenza virus in HEK-293 cells for efficient vaccine manufacturing* Vaccine, **28**, 3661–3671

Makarkov, A.I., Patel, A.R., Bainov, V. and Ward, B.J. (2018) *A novel serological assay for influenza based on DiD fluorescence dequenching that is free from observer bias and potentially automatable - A proof of concept study* Vaccine, **36**, 4485–4493

Matassov, D., Cupo, A. and Galarza, J.M. (2007) *A novel intranasal virus-like particle (VLP) vaccine designed to protect against the pandemic 1918 influenza A virus (H1N1)* Viral Immunol., **20**, 441-452

Shaw, M.L., Stone, K.L., Colangelo, C.M., Gilcicek, E.E. and Palese, P. (2008) *Cellular proteins in influenza virus particles* PLoS Pathog., **4**:e1000085

Smith, T., O'Kennedy, M.M., Wandrag, D.B.R., Adeyemi, M. and Abolnik, C. (2020) *Efficacy of a plant-produced virus-like particle vaccine in chickens challenged with Influenza A H6N2 virus* Plant Biotech. J., **18**, 502–512

Speshock, J.L., Doyon-Reale, N., Rabah, R., Neely, M.N. and Roberts, P.C. (2007) *Filamentous influenza virus A infection predisposes mice to fatal septicemia following superinfection with Streptococcus pneumoniae Serotype 3* Infect. Immun., **75**, 3102-3111

Su, W-C., Yu, W-Y., Huang, S-H. and Laia, M.M.C. (2018) *Ubiquitination of the cytoplasmic domain of influenza A virus M2 protein is crucial for production of infectious virus particles* J. Virol., **92**: e01972-17

Sulli, C., Banik, S.S.R., Schilling, J., Moser, A., Xiang, X., Payne, R., Wanless, A., Willis, S.H., Paes, C., Rucker, J.B. and Doranz, B.J. (2013) *Detection of proton movement directly across viral membranes to identify novel influenza virus M2 inhibitors* J. Virol., **87**, 10679-10686

Thompson, C.M., Petiot, E., Lennaertz, A., Henry, O. and Kamen, A.A. (2013) *Analytical technologies for influenza virus-like particle candidate vaccines: challenges and emerging approaches* Virol. J. **10**: 141

Thompson, C.M., Petiot, E., Mullick, A., Aucoin, M.G., Henry, O. and Kamen, A.A. (2015) *Critical assessment of influenza VLP production in Sf9 and HEK293 expression systems* BMC Biotechnol., **15**: 31

Yang, X., Steukers, L., Forier, K., Xiong, R., Braeckmans, K., Van Reeth, K. and Nauwynck, H. (2014) *A beneficiary role for neuraminidase in influenza virus penetration through the respiratory mucus* PLoS One, **9**: e110026
Yang, Y., Leggat, D., Herbert, A., Roberts, P.C. and Sundick, R.S. (2009) *A novel method to incorporate bioactive cytokines as adjuvants on the surface of virus particles* J. Interferon Cytokine Res., **29**, 9-23

6 *Paramyxoviridae*
Henipavirus
Akiyama, H., Miller, C., Patel, H.V., Hatch, S.C., Archer, J., Ramirez, N-G.P. and Gummuluru, S. (2014) *Virus particle release from glycosphingolipid-enriched microdomains is essential for dendritic cell-mediated capture and transfer of HIV-1 and Henipavirus* J. Virol., **88**, 8813–8825

Paramyxovirinae
Avlulavirus
Newcastle disease virus
Biswas, M., Johnson, J.B., Kumar, S.R.P., Parks, G.D. and Subbiaha, E. (2012) *Incorporation of host complement regulatory proteins into Newcastle disease virus enhances complement evasion* J. Virol., **86**, 12708-12716

Morbillivirus
Measles virus
Brindley, M.A. and Plemper, R.K. (2010) *Blue native PAGE and biomolecular complementation reveal a tetrameric or higher-order oligomer organization of the physiological measles virus attachment protein H* J. Virol., **84**, 12174-12184
Liljeroos, L., Huiskonen, J.T., Ora, A., Susi, P. and Butcher, S.J. (2011) *Electron cryotomography of measles virus reveals how matrix protein coats the ribonucleocapsid within intact virions* Proc. Natl. Acad. Sci. USA, **108**, 18085–18090
Hallak, L.K., Merchan, J.R., Storgard, C.M., Loftus, J.C. and Russell, S.J. (2005) *Targeted measles virus vector displaying echistatin infects endothelial cells via* $\alpha v\beta 3$ *and leads to tumor regression* Cancer Res., **65**, 5292-5300

Pneumovirinae
Human respiratory syncytial virus
Gias, E., Nielsen, S.U., Morgan, L.A.F. and Toms, G.L. (2008) *Purification of human respiratory syncytial virus by ultracentrifugation in iodixanol density gradient* J. Virol. Methods, **147**, 328-332
Murawski, M.R., Bowen, G.N., Cerny, A.M., Anderson, L.J., Haynes, L.M., Tripp, R.A., Kurt-Jones, E.A. and Finberg, R.W. (2009) *Respiratory syncytial virus activates innate immunity through Toll-Like receptor 2* J. Virol., **83**, 1492-1500

Respirovirus
Swine paramyxovirus
Qiao, D., Janke, B.H. and Elankumaran, S. (2009) *Molecular characterization of glycoprotein genes and phylogenetic analysis of two swine paramyxoviruses isolated from United States* Virus Genes, **39**, 53–65

6 *Rhabdoviridae*
Lyssavirus
Rabies virus
Chatterjee, S., Sullivan, H.A., MacLennan, B.J., Xu, R., Hou, Y.Y., Lavin, T.K., Lea, N.E., Michalski, J.E., Babcock, K.R. et al (2018) *Nontoxic, double-deletion-mutant rabies viral vectors for retrograde targeting of projection neurons* Nat. Neurosci., **638**, 638–646
Finke, S. and Conzelmann, K-K. (2003) *Dissociation of rabies virus matrix protein functions in regulation of viral RNA synthesis and virus assembly* J. Virol., **77**, 12704-12082
Finke, S., Brzozka, K. and Conzelmann, K-K. (2004) *Tracking fluorescence-labeled rabies virus: enhanced green fluorescent protein-tagged phosphoprotein P supports virus gene expression and formation of infectious particles* J. Virol., **78**, 12333-12343
Fontana, D., Kratje, R., Etcheverrigaray, M. and Prieto, C. (2015) *Immunogenic virus-like particles continuously expressed inmammalian cells as a veterinary rabies vaccine candidate* Vaccine, **33**, 4238–4246
Fontana, D., Etcheverrigaray, M., Kratje, R. and Prieto, C. (2016) *Development of rabies virus-like particles for vaccine applications: production, characterization, and protection studies* In Vaccine Design: Methods and

Protocols, Vol. 1: Vaccines for Human Diseases, Methods in Molecular Biology, vol. 1403 (ed. Thomas, S.) Springer Science+Business Media New York pp 155-166
Fontana, D., Marsili, F., Garay, E., Battagliotti, J., Etcheverrigaray, M., Kratje, R. and Prieto, C. (2019) *A simplified roller bottle platform for the production of a new generation VLPs rabies vaccine for veterinary applications* Comp. Immunol. Microbiol. Infect. Dis., **65**, 70–75
Hidaka, Y., Lim, C-K., Takayama-Ito, M., Park, C-H., Kimitsuki, K., Shiwa, N., Inoue, K-i., Itou, T. (2018) *Segmentation of the rabies virus genome* Vir. Res., **252**, 68–75
Klingen, Y., Conzelmann, K-K. and Finke, S. (2008) *Double-labeled rabies virus: live tracking of enveloped virus transport* J. Virol., **82**, 237-245
Marschalek A., Drechsel, L. amd Conzelmann, K-K. (2012) *The importance of being short: The role of rabies virus phosphoprotein isoforms assessed by differential IRES translation initiation* Eur. J. Cell Biol., **91**, 17– 23

Vesiculovirus
Vesicular stomatitis virus

Andreu-Moreno, I. and Sanjuán, R. (2018) *Collective infection of cells by viral aggregates promotes early viral proliferation and reveals a cellular-level Allee effect* Curr, Biol., **28**, 3212–3219
Arulanandam, R., Batenchuk, C., Varette, O., Zakaria, C., Garcia, V., Forbes, N.E., Davis, C. Krishnan, R. et al (2015) *Microtubule disruption synergizes with oncolytic virotherapy by inhibiting interferon translation and potentiating bystander killing* Nat. Commun., **6**: 6410
Betancourt, D., Ramos, J.C. and Barber, G.N. (2015) *Retargeting oncolytic vesicular stomatitis virus to human T-cell lymphotropic virus Type 1-associated adult T-cell leukemia* J. Virol., **89**, 11786-11800
Betancourt, D., de Queiroz, N.M.G.P., Xia, T., Ahn, J. and Barber, G.N. (2017) *Cutting edge: innate immune augmenting vesicular stomatitis virus expressing Zika virus proteins confers protective immunity* J. Immunol., **198**, 3023–3028
Beug, S.T., Beauregard, C.E., Healy, C., Sanda, T., St-Jean, M., Chabot, J., Walker, D.E., Mohan, A., Earl, N. et al (2017) *Smac mimetics synergize with immune checkpoint inhibitors to promote tumour immunity against glioblastoma* Nat. Comm., **8**: 14278
Beug, S.T., Pichette, S.J., St-Jean, M., Holbrook, J., Walker, D.E., LaCasse, E.C. and Korneluk, R.G. (2018) *Combination of IAP antagonists and TNF-α-armed oncolytic viruses induce tumor vascular shutdown and tumor regression* Mol. Ther: Oncolytics, **10**, 28-39
Cuevas, J.M., Durán-Moreno, M. and Sanjuán, R. (2017) *Multi-virion infectious units arise from free viral particles in an enveloped virus* Nat. Microbiol., **2**: 17078
Diallo, J-S., Le Boeuf, F., Lai, F., Cox, J., Vaha-Koskela, M., Abdelbary, H., MacTavish, H., Waite, K., Falls, T. et al (2010) *A high-throughput pharmacoviral approach identifies novel oncolytic virus sensitizers* Mol. Ther., **18**, 1123-1129
Diallo, J-S., Vähä-Koskela, M., Le Boeuf, F. and Bell, J. (2011) *Propagation, purification, and in vivo testing of oncolytic vesicular stomatitis virus strains* In Methods Mol. Biol., **797,** Oncolytic Viruses: Methods and Protocols, (ed. Kirn, D.H. et al.), Springer Science+Business Media, pp 127-140
Domingo-Calap, P., Segredo-Otero, E., Durán-Moreno, M. and Sanjuán, R. (2019) *Social evolution of innate immunity evasion in a virus* Nat. Microbiol., **4**, 1006–1013
Dornan, M.H., Krishnan, R., Macklin, A.M., Selman, M., El Sayes, N., Son, H.H., Davis, C., Chen, A., Keillor, K., Le, P.J. et al (2016) *First-in-class small molecule potentiators of cancer virotherapy* Sci. Rep., **6**: 26786
Garijo, R., Hernández-Alonso, P., Rivas, C., Diallo, J-S. and Sanjuán, R. (2014) *Experimental evolution of an oncolytic vesicular stomatitis virus with increased selectivity for p53-deficient cells* PLoS One, **9**: e102365
Gélinas, J-F., Azizi, H., Kiesslich, S., Lanthier, S., Perdersen, J., Chahal, P.S., Ansorge, S., Kobinger, G. et al (2019) *Production of rVSV-ZEBOV in serum-free suspension culture of HEK 293SF cells* Vaccine, **37**, 6624–6632
Hastie, E., Besmer, D.M., Shah, N.R., Murphy, A.M., Moerdyk-Schauwecker, M., Molestina, C., Das Roy, L., Curry, J.M., Mukherjee, P. and Grdzelishvili, V.Z. (2013) *Oncolytic vesicular stomatitis virus in an immunocompetent model of MUC1-positive or MUC1-null pancreatic ductal adenocarcinoma* J. Virol., 87, 10283–10294
Kalvodova, L., Sampaio, J.L., Cordo, S., Ejsing, C.S., Shevchenko, A. and Simons, K. (2009) *The lipidomes of vesicular stomatitis virus, Semliki Forest virus and the host plasma membrane analyzed by quantitative shotgun mass spectrometry* J. Virol., **83**, 7996-8003
Kim, D-S., Dastidar, H., Zhang, C., Zemp, F.J., Lau, K., Ernst, M., Rakic, A., Sikdar, S., Rajwani, J. et al (2017) *Smac mimetics and oncolytic viruses synergize in driving anticancer T-cell responses through complementary mechanisms* Nat. Comm., **8**: 344
Moerdyk-Schauwecker, M,. Hwang, S-I., Grdzelishvili, V.Z. (2014) *Cellular proteins associated with the interior and exterior of vesicular stomatitis virus virions*. PLoS One, **9**: e104688

Rodriguez, S.E., Cross, R.W., Fenton, K.A., Bente, D.A., Mire, C.E. and Geisbert, T.W. (2019) *Vesicular stomatitis virus-based vaccine protects mice against Crimean-Congo hemorrhagic fever* Sci. Rep., 9: 7755
Selman, M., Ou, P., Rousso, C., Bergeron, A., Krishnan, R., Pikor, L., Chen, A., Keller, B.A., Ilkow, C., Bell, J.C. and Diallo, J-S. (2018) *Dimethyl fumarate potentiates oncolytic virotherapy through NF-κB inhibition* Sci. Transl. Med., **10**: eaao1613

OptiPrep™ Reference List RV05: 3rd edition, January 2020